

Artificial Insemination Centres' Health Management Program

(Programme de gestion sanitaire
des centres d'insémination artificielle) - PGSCIA

2023-2024

Registration Conditions



Centre de développement
du porc du Québec inc.

©Centre de développement du porc du Québec inc.
Legal deposit - 2023
Bibliothèque et Archives nationales du Québec
Library and Archives Canada
ISBN 978-2-925175-20-9
ISSN 2291-3572

Table of Content

1	PGSCIA Eligibility Terms and Conditions	2
1.1	Registration.....	2
1.2	Preconditions	2
1.3	Termination of services and re-registration	2
2	Respective responsibilities of the three stakeholders	3
2.1	Responsibilities of the CDPQ-designated veterinarian.....	3
2.2	Responsibilities of insemination centre managers.....	3
2.3	Responsibilities of the insemination centre’s consultant veterinarian	4
3	Biosecurity of centres.....	5
3.1	Basic biosecurity guidelines	5
4	Minimum standards for PGSCIA AI member centres.....	7
4.1	Isolation (quarantine) premises.....	7
4.2	Boar stud premises	7
4.3	Semen packaging laboratory	8
4.4	Semen storage and shipping room	8
4.5	Drug cabinet.....	8
5	Management of farm personnel and equipment	9
5.1	Farm personnel management	9
5.2	Managing Equipment and Supplies	9
6	Selection of batches of animals for admission to AI centres	10
6.1	Location of supplier farms	10
6.2	Boars from a PRRS virus-free farm site (PRRS-naïve status)	10
6.3	Boars from a farm where PRRS virus has been eradicated (PRRS-negative status)	11
7	List of diseases under surveillance	12
7.1	Surveillance program	12
7.2	Treatment and vaccination program (quarantine and boar units)	13
8	Macro plan of the testing strategy.....	14
8.1	Blood draw(s) A.....	14
8.2	Blood draw B.....	15
8.3	Blood draw C.....	15

8.4	Blood draw D.....	15
8.5	Blood draw E	15
8.6	Blood draws F-Z	16
9	Health status certification for boars destined for AI centres (PRRS)	17
9.1	Recommended laboratory tests	17
9.2	Evaluation Strategy	17
9.3	Interpretation of the ELISA-X3 test on serum	17
9.4	Interpretation of the PCR test	17
9.5	Possible responses	18
10	Health status certification for boars destined for AI centres (APP)	19
10.1	Recommended laboratory test.....	19
10.2	Evaluation Strategy	19
10.3	Interpretation of the ELISA-LC-LPS test	19
10.4	Possible results	19
11	Health status certificate for boars destined for centres (PEDV, TGE and SDCV).....	20
11.1	Recommended laboratory test.....	20
11.2	Evaluation Strategy	20
11.3	Interpretation of the PCR test	20
11.4	Possible results	21
12	Health status certificate for boars destined for AI centres (PCVAD)	22
12.1	Porcine Circovirus Associated Diseases (PCVAD) – diagnostic criteria for infection among a herd.....	22
12.2	Evaluation Strategy	22
13	Certification of the health status of boars for the centres (Senecavirus A)	23
13.1	Recommended laboratory technique.....	23
13.2	Evaluation strategy	23
13.3	Interpretation of the PCR tests.....	23
13.4	Possible answers	24
14	Surveillance procedures for boar stud units (PRRS)	25
14.1	Recommended laboratory tests	25
14.2	Minimum testing strategy by boar population.....	25
14.3	Possible responses	25
14.4	Monitoring the incidence of health problems potentially linked to PRRS	26

15	Emergency procedures in case of a PRRS alert in an AIC.....	27
15.1	Positive blood test result	27
15.2	Increase in health problems	27
15.3	Possible responses	27
16	Procedures to follow whenever an AIC is infected with PRRS virus	28
16.1	Insemination centre is positive.....	28
17	Surveillance procedures for boar stud units (PEDV, TGE and SDCV)	29
17.1	Recommended laboratory tests	29
17.2	Minimum testing strategy by boar population.....	29
17.3	Possible responses	29
17.4	Monitoring the incidence of health problems potentially linked to PEDV and SDCV.....	29
18	Emergency procedures in case of a PEDV, TGE or SDCV alert in an AIC	30
18.1	Positive blood test result	30
18.2	Increase in health problems	30
18.3	Possible responses	30
19	Procedures to follow when an AIC is infected with by PEDV, TGE or SDCV virus.....	31
19.1	Positive insemination centre	31
20	Microbiological quality control of the semen	32
20.1	Objectives	32
20.2	Methods.....	32
	Appendix 1 - Registration Form for AI centres	I
	Appendix 2 - Contact Details for the Resource Persons	II
	Appendix 3 - CDPQ Health Certificate for Boar Entry into the AIC (D03).....	III
	Appendix 4 - Laboratories.....	IV
	Appendix 5 – Sampling guide for boar purchases (D02)	v

Introduction

The *Programme de gestion sanitaire des centres d'insémination artificielle* (PGSCIA) is a health management program for artificial insemination centres, open to all AI centres wishing to sell boar semen to Quebec farmers and hog producers.

PGSCIA is an adjunct to the program administered by the Canadian Food Inspection Agency (CFIA).

The main objectives are to:

- ❖ Attest to the quality of each insemination centre's health management program to monitor the procedures for verifying the safety of boar semen sold to the user farms;
- ❖ Check that the AI centres meet minimum standards for facilities and equipment, hygiene, health and safety, record keeping, semen traceability and animal health control;
- ❖ Provide AI centre managers with the tools and expertise to improve centre biosecurity regarding PRRS virus, PED, SDCV, TGE, SVA contamination;
- ❖ Provide centre managers with tools and procedures to lower the risk of the semen spreading potentially harmful viral pathogens to the user farms;
- ❖ Certify the health status of boars purchased by the insemination centres to minimize the risk of animal health accidents; and
- ❖ Put forward and improve health status verification procedures for the insemination centres to attest to their health status regarding PRRS virus, PED, DCVP, TGE, SVA.

1 PGSCIA Eligibility Terms and Conditions

1.1 Registration

Fill out the PGSCIA Registration Form for AI Centres (Appendix 1) and send it to the CDPQ (see Appendix 2).

1.2 Preconditions

To qualify for admission to the PGSCIA program an AI centre:

- a. Must be a registered farm business with the Bureau de renseignements agricoles (agricultural information office) of MAPAQ, in accordance with provincial regulations;
- b. Must be a Canadian Food Inspection Agency approved semen production and distribution centre in accordance with the Health of Animals Act and Regulations (L.C.1990, ch. 21);
- c. Must comply with the standards set out in the program regarding facilities and equipment, hygiene and health protection (biosecurity), record keeping and control of animal health;
- d. Must use the services of a consultant veterinarian responsible for the development, maintenance and implementation of the health management program; and
- e. Must be officially recognized by the Canadian Quality Assurance Program (CQA) and the Animal Care Assessment (ACA) of the Canadian Pork Council (CPC).

1.3 Termination of services and re-registration

Insemination centre members that no longer meet the program requirements in force on April 1, 2023, together with those centres that do not conform, can be suspended or expelled at any time.

Member centres are automatically re-registered on April 1st of each year, unless otherwise specified in writing.

Any centre wishing to abandon the program must provide prior written notice to the CDPQ.

2 Respective responsibilities of the three stakeholders

2.1 Responsibilities of the CDPQ-designated veterinarian

- Develop, enhance, adapt the program to the needs of Quebec's pork industry.
- Inspect the member AI centre's facilities once a year. The purpose of these visits is to verify that the standards for facilities, hygiene, health and safety, record keeping and control of animal health, are applied in accordance with the insemination centre's health management program.
- Authorize animal admission to the isolation (quarantine) unit and, at the client's request, authorize admission of boars to the boar unit/barn.
- Maintain regular contact with the AI centre manager, the centre's consultant veterinarian and the Canadian Food Inspection Agency veterinarian responsible for the program.

2.2 Responsibilities of insemination centre managers

- Comply with the requirements for admission to the program.
- Promptly notify the consultant veterinarian and the veterinarian responsible for PGSCIA (hereafter, the CDPQ-designated veterinarian) in the event of potentially contagious health problems.
- Authorize access to AI centre facilities by the CDPQ-designated veterinarian at all times.
- Provide the CDPQ-designated veterinarian with all health information concerning the AI centre that relates to the program.
- Allow the CDPQ-designated veterinarian to consult AI centre records related to the program.
- Implement health recommendations from the CDPQ-designated veterinarian and the consultant veterinarian without delay.
- Not distribute any semen that, insofar as they know, would present a health risk for the AI centre's clients.

2.3 Responsibilities of the insemination centre's consultant veterinarian

- Inspect centre facilities every four months (minimum three times per year). At least one of these visits will be conducted together with the CDPQ-designated veterinarian.
- During these visits, verify that the standards for facilities, hygiene, biosecurity, record keeping, and animal health control are being applied.
- Following these visits, write recommendations for the AI centre manager and the CDPQ-designated veterinarian concerning facilities, sanitation, biosecurity, record keeping and animal health control.
- In the event of potentially contagious health problems, recommend temporary suspension of the semen distribution and immediately notify the CDPQ-designated veterinarian.
- Write the necessary prescriptions for the purchase of drugs or vaccines and ensure they are used correctly.
- Prescribe the necessary laboratory tests for managing the health of the boar stud.
- Collaborate in maintaining and updating the insemination centre's health management program.

3 Biosecurity of centres

3.1 Basic biosecurity guidelines

- ❖ Identification of critical points/issues
- ❖ Development of an animal health management program for each AI centre
- ❖ Regular verification of compliance with program guidelines

3.1.1 Identification of critical points

- Location of the premises
- Management of farm personnel and equipment
- Selection of livestock suppliers
- Health status certificate for boars entering the AI centre
- Bacteriological quality of the semen
- Transport of animals:
 - From farm to isolation (quarantine) unit, from isolation (quarantine) unit to boar stud unit, from boar stud unit to slaughterhouse
- Cleaning and disinfection of the trucks
- Transportation of the semen
- Monitoring of vehicle movement around the premises
- Vermin and pest control (birds, rodents, flies, etc.)

3.1.2 Development of an animal health management program for each AI centre

Each member AI centre of PGSCIA must have a written document outlining in detail its health management program. This document must be updated annually on October 1 each year.

The CDPQ-designated veterinarian may participate in the development of the insemination centre's health management program. However, maintenance of the program and updates to it are the responsibility of AI centre managers and of their consultant veterinarian.

CDPQ can provide an example of a health management program to help orient AI centre managers.

3.1.3 Regular verification of compliance with program guidelines

Centre managers must check the application of the programs (regularly); the consultant veterinarian must do so as well (at least three times a year) and the CDPQ-designated veterinarian also (once a year).

4 Minimum standards for PGSCIA AI member centres

4.1 Isolation (quarantine) premises

- Must be located more than 1 km from swine operations areas other than artificial insemination centres (AIC);
- Ideally, the isolation (quarantine) units for AICs that are not equipped with an air filtration system at the entrance must be located at a distance of more than one kilometer from the AIC premises;
- Must be accessible to authorized farm personnel only (locked doors);
- Must be equipped with a Danish entry or a shower at the entrance and must follow the principles of unidirectional flow;
- Must be designed so as to prevent the entry of vermin, birds or other animals;
- Must be regularly inspected for flies and vermin by a professional exterminator;
- Must be operated on all-in/all-out (AI/AO) basis per unit;
- Must be sufficiently spacious, airy and comfortable to guarantee the welfare of the animals; and
- Must be cleaned and disinfected and then kept empty for at least five days between each occupation.

4.2 Boar stud premises

- Must be located at a minimum distance of 1 km from other area groups of pigs other than the AIC's isolation (quarantine) unit;
- Must be separated by a physical barrier from other unit areas of the AI centre;
- Must be accessible to authorized farm personnel only (locked doors);
- Must be equipped with a Danish entry or a shower at the entrance and must follow the principles of unidirectional movement;
- Must be designed so as to prevent entry of vermin, birds or other animals;
- Must be regularly inspected for flies and vermin by a professional exterminator; and
- Must be sufficiently spacious, airy and comfortable to guarantee the welfare of the animals.

4.3 Semen packaging laboratory

- Must be separated by a physical barrier from other unit areas of the centre;
- Must be accessible to authorized persons only;
- Must be separated from the collection area by an airlock to prevent airborne contamination; and
- Shall not permit any backward people flow from the other unit areas.

4.4 Semen storage and shipping room

- Must be separated by a physical barrier from other unit areas of the AI centre; and
- Must be accessible to authorized persons only.

4.5 Drug cabinet

- Must be designed to allow the storage of drugs and vaccines in accordance with the manufacturer's recommendations and the drug regulation requirements (e.g. CQA).

5 Management of farm personnel and equipment

5.1 Farm personnel management

- Entrance doors to boar and quarantine units must be locked at all times.
- All farm personnel and visitors must change street clothes and shoes before entering the boar stud units.
- Staff and visitors must observe a 48-hour downtime period (two nights) without any direct contact with pigs before entering boar stud premises.
- International visitors who have had no contact with pigs must observe a 2 to 7 days downtime period depending of the country.
- International visitors who have been in contact with pigs must observe a 2 to 7 days downtime period depending of the country.
- All visitors must sign the visitor log book upon entering a boar stud premises.
- Visitors and farm personnel must remove all jewelry before entering the boar stud premises.

5.2 Managing Equipment and Supplies

- All equipment and supplies that come into the boar stud units and quarantine units must be new and disinfected.
- A workroom where fumigation of equipment and supplies can take place is strongly recommended.

6 Selection of batches of animals for admission to AI centres

6.1 Location of supplier farms

- Any breeding operation with no pig site (pig farm, slaughterhouse, assembly station) within 5 km meets the location standards as a boar supplier.
- Any farm with no pig production site within a 2 km radius and an exposure index¹ of less than 2 could qualify.

¹ *The exposure index is an estimate of the exposure of a boar-supplying site in relation to sites with pigs in the area. Sites with an epidemiological link are not included in the estimation.*

6.2 Boars from a PRRS virus-free farm site (PRRS-naïve status)

6.2.1 Characteristics of the Maternity unit

- No clinical signs of PRRS
- Neonatal mortality (birth to weaning) is well controlled with an objective of less than 18%

6.2.2 Characteristics of the Nursery and Grow-to-Finish units

1. Death losses (monthly mortality rate) are:
 - less than 5% in nursery unit.
 - less than 5% in finishing unit.
 - less than 8-10% between weaning and the end of finishing.
2. Documented seronegative results for PRRS using regular (a) or irregular (b) serological monitoring:
 - a. Regular serological monitoring (> 70 kg): minimum of 10 negative subjects per month (ELISA-X3 or other);
 - b. Irregular serological monitoring (> 70 kg): minimum of 20 negative subjects for six months (ELISA-X3 or other)
3. Absence of clinical signs consistent with porcine circovirus associated diseases (PCVAD) (see p. 24, [Health certificate – PCVAD](#))
4. No clinical signs of other diseases under surveillance (see [List of diseases under surveillance](#)).

5. Obtaining laboratory results (see pp.18-19) for the selected boars within the 30 days preceding entry into isolation (quarantine) unit.

6.3 Boars from a farm where PRRS virus has been eradicated (PRRS-negative status)

6.3.1 Characteristics of Maternity unit (first year)

- Absence of clinical signs of PRRS; and
- Neonatal mortality (farrow-to-wean) well controlled with an objective of less than 18%.

6.3.2 Characteristics of Maternity unit (subsequent years)

- Same as for PRRS-free farm (PRRS-naïve status).

6.3.3 Characteristics of the nursery and grow finish

1. Death losses (monthly mortality rate) are:
 - less than 5% in nursery unit.
 - less than 5% in finishing unit.
 - less than 8-10 % between weaning and the end of finishing.
2. Documented seronegative results for PRRS using regular (A) or irregular (B) serological monitoring:
 - a. Regular serological monitoring (> 70 kg): minimum of 10 negative subjects per month (ELISA-X3 or other);
 - b. Regular serological monitoring (> 70 kg): minimum of 20 negative subjects for six months (ELISA-X3 or other)
3. Absence of clinical signs consistent with porcine circovirus associated diseases (PCVAD) (see p. 24, [Health certificate – PCVAD](#))
4. No clinical signs of other diseases under surveillance (see [List of diseases under surveillance](#)).
5. Obtaining laboratory results (see pp.18-19) for the selected boars within the 30 days preceding entry into isolation (quarantine) unit.
 1. Seronegative (ELISA-X3) and PCR-negative for PRRS, within one full week preceding transport to the isolation (quarantine) unit. Blood sampling procedures on the selected animals must be done in accordance with the procedures of the PGSCIA program (pp.15-16-17).

7 List of diseases under surveillance

7.1 Surveillance program

- ❖ Animals in the source herds should display no clinical signs of:
 - Enzootic pneumonia
 - Porcine pleuropneumonia (*Actinobacillus pleuropneumoniae*)
 - *Actinobacillus suis*
 - Glässer's disease (*Glaesserella parasuis*)
 - Porcine reproductive and respiratory syndrome (PRRS)
 - Proliferative enteropathy/ Porcine proliferative enteritis
 - Porcine Circovirus Associated Diseases (PCVAD)
 - Dysentery
 - Atrophic rhinitis
 - Sarcoptic mange
 - Porcine epidemic diarrhea (PEDV)
 - Delta coronavirus (SDCV),
 - Transmissible gastroenteritis (TGE)
 - Senecavirus A
 - Influenza
 - Salmonellosis
 - Leptospirosis

- ❖ Selected boars must be free of:
 - Porcine reproductive and respiratory syndrome (PRRS)
 - Porcine pleuropneumonia *Actinobacillus pleuropneumoniae* (App serotypes 1 and 5)
 - Transmissible gastroenteritis (TGE)
 - Porcine epidemic diarrhea (PEDV), and
 - Delta coronavirus (SDCV)
 - Senecavirus A

7.2 Treatment and vaccination program (quarantine and boar units)

7.2.1 Sarcoptic mange

Treatment and eradication of sarcoptic mange (*Sarcoptes scabiei*) must be carried out in the isolation (quarantine) unit, when the source is considered positive.

7.2.2 Porcine circovirus

It is strongly recommended that boars have been vaccinated on at least two occasions in their lives against porcine circovirus before entering the boar stud unit:

- a single vaccination in the isolation (quarantine) unit is recommended for boars that have already been vaccinated at the farm of origin.
- double vaccination in the isolation (quarantine) unit is recommended for boars who have never been vaccinated against porcine circovirus.

7.2.3 Glässer's disease (*Glaesserella parasuis*) and *Mycoplasma Hyopneumoniae*

Vaccination in the isolation (quarantine) unit is especially recommended for boars from disinfected herds.

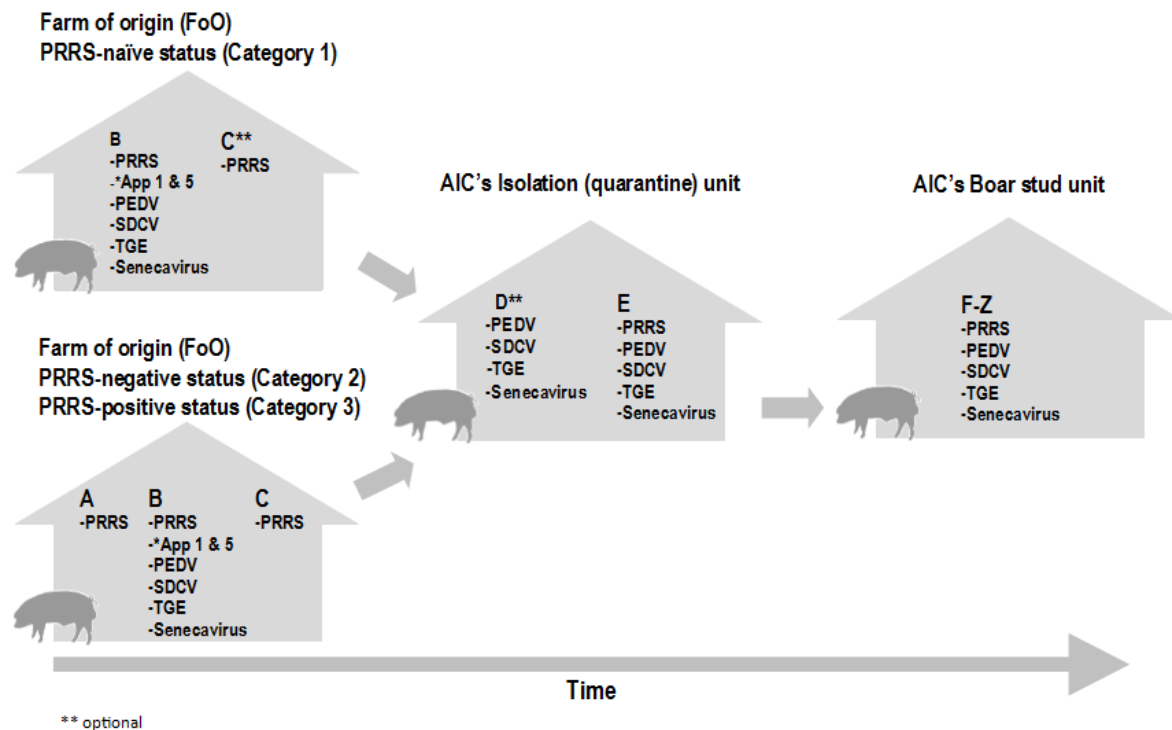
7.2.4 Porcine parvovirus and Swine erysipelas (*Erysipelothrix rhusiopathiae*).

Animals in the boar units must be vaccinated twice a year.

7.2.5 Endoparasites

Treatment against internal parasites (endoparasites) should be carried out every six months in the boar units. When no treatment is done in quarantine for mange, then the boars need to be treated for internal parasites.

8 Macro plan of the testing strategy



The macro plan shows the blood tests performed on boars from the farm of origin to the boar stud unit. PRRS virus, App serotypes 1 and 5, PEDV, TGE, SDCV and SVA status is checked by the CDPQ program.

* Serology for App serotypes 1 and 5 is required for consignments sourced from farms that have a history of infection, or a history of seropositive results, or that are of unknown status).

8.1 Blood draw(s) A

One or more blood draws can be done during the isolation (quarantine) period according to instructions from the CDPQ-designated veterinarian. Blood serums can be tested for PRRS using ELISA-X3 and PCR and if needed, by MFIA or Biocheck.

8.2 Blood draw B

This blood draw is done by a CFIA-accredited veterinarian on the farm of origin or in the farm of origin isolation (quarantine) unit.

- **SRRP and App:** Serums or oral fluids are sent for PRRS serological testing (Appendix 6) and, if necessary, serums for App serotypes 1 and 5 testing.
- **PED, SDCV, TGE and Senecavirus A:** oral fluid samples taken with the rope technic are done in a minimum of 4 pens; alternatively, manure samples are taken in a minimum of 4 pens with the possibility to sample the boots of the producer or the walkway. The samples can be tested in pools (1 pool of 4) at the laboratory before running the PCR tests.

8.3 Blood draw C

This blood draw is optional for farms with PRRS-naïve status and is mandatory for PRRS-negative and PRRS-positive status farms. The CDPQ-designated veterinarian can insist on a blood draw and further testing within 7 days before the animals are transported to the AIC's isolation (quarantine) unit. Serums can be tested for PRRS using ELISA-X3 and PCR and if needed, by MFIA or Biocheck.

8.4 Blood draw D

This blood draw is optional and is done 3 to 5 days after the start of quarantine by the quarantine personnel. A pool of 5 fecal samples per building will be taken to perform the PCR tests for PEDV, SDCV, TGE and Senecavirus A.

8.5 Blood draw E

This blood draw is done by staff from the insemination centre three to five days before the transport to the boar stud unit.

- ❖ PRRS:
 - The serums are verified by a serological test for all boars;
- ❖ PEDV, SDCV, GET and Senecavirus A:
 - A pool of 5 fecal samples per building will be taken to perform the PCR tests for PEDV, SDCV, TGE and Senecavirus A.

8.6 Blood draws F-Z

These blood draws are done at least once a week to verify the status of each population of boars:

- ❖ PRRS: a weekly PCR on a pool of ten serums and monthly ELISA-X3 tests on a minimum of five serums.
- ❖ PEDV, SDCV, GET and Senecavirus A: weekly PCR testing on a pool of 5 fecal samples.

9.5 Possible responses

Serum

- ❖ All results are negative for ELISA-X3 and PCR tests:
 - All boars are accepted.
- ❖ Certain boars are suspect or positive with the ELISA-X3 test and negative with the PCR test.
 - Complementary test with MFIA or Biocheck test on all serum from suspected and positive ELISA-X3 cases (5 serum in total with higher S/Ps)
 - MFIA or Biocheck test are negative:
 - The batch is declared negative; boars with negative status with ELISA-X3 test are accepted; boars with positive or suspect status with ELISA-X3 test are refused.
 - MFIA or Biocheck test positive:
 - Several boars have a suspect or positive result relating to the ELISA-X3 test and MFIA or Biocheck test:
 - If it is only an individual, the boar and nine other boars in contact with it are individually retested 10 days later with an ELISA-X3 test and MFIA or Biocheck test. A PCR PRRS test in pools of 5 is also required.
 - Only the individual that tested positive has remained positive after the second ELISA-X3 and MFIA or Biocheck test, all PCR are negative:
 - ✓ the batch is declared negative: the negative-status boars relating to the ELISA-X3 test are accepted; the boars that are positive or suspect are refused.
 - The individual that previously tested positive and one additional boar is now positive after the second ELISA-X3 test and MFIA or Biocheck test, and the PCRs are all negative:
 - ✓ the veterinarian designated by the CDPQ demands a new blood testing of the boars and the entire assessment process is restarted.
 - The individual that previously tested positive and one additional boar is now positive after the second ELISA-X3 test and MFIA or Biocheck test and at least one of the pool PCR is positive:
 - ✓ all the animals are rejected.
 - If two or more individuals test positive, the veterinarian designated by the CDPQ demands a new blood testing of the boars and the evaluation process is restarted.

11 Health status certificate for boars destined for centres (PEDV, TGE and SDCV)

11.1 Recommended laboratory test

- ❖ Test PEDV-TGE-SDCV PCR from Tetracore (to detect virus);
- ❖ Test PEDV PCR, TGE PCR and SDCV PCR of the Faculté de médecine vétérinaire de l'Université de Montréal (to detect virus).

11.2 Evaluation Strategy

- ❖ A signed health certificate confirming the absence of clinical signs of disease throughout the growth period received within **two weeks** prior to moving the boars from the farm to AI centres quarantine
- ❖ Verify the absence of the virus with the PCR technique using one of two options:
 - ❖ **Option A)** Oral fluids with the rope technique in a minimum of four pens;
 - ❖ **Option B)** Manure samples are taken in a minimum of 4 pens with the possibility to sample the boots of the producer or the walkway.
 - The samples can be pooled (maximum of 4) at the laboratory before testing for the virus with PCR tests.
- ❖ The sampling should be done as close as possible to the shipment of the boars and no more than 30 days prior shipment of the boars in quarantine.

11.3 Interpretation of the PCR test

Each sample is interpreted individually:

- ❖ Interpretation of the PEDV-TGE-SDCV PCR test from Tetracore:
 - $CT \leq 35$ ➡ positive sample;
 - $CT > 35$ et ≤ 38 ➡ suspect sample;
 - $CT > 38$ ➡ negative sample.
- ❖ Interpretation of the PEDV-TGE-SDCV PCR test from the Faculté de médecine vétérinaire:
 - $CT \leq 35$ ➡ positive sample;

- CT > 35 et ≤ 40 ➡ suspect sample;
- CT > 40 ➡ negative sample.

Note: The interpretation of these tests can change according new scientific knowledge which will emerge

11.4 Possible results

- ❖ All results in the PEDV-TEG-SDCV PCR test are negative:
 - All the boars are accepted.

- ❖ Some boars are suspect or positive:
 - Ask for additional testing on positive samples already in the laboratory:
 - Separate (one-pool) and run again the PCR test on each sample.
 - Take another set of oral fluids (minimum of 4) or manure samples in 4 pens (the ones that are around the first set of samples):
 - Run the PCR test on oral fluids or on manure samples.

If at least one sample turns out positive for one of these tests, no boars will be admitted to the AI centre.

12 Health status certificate for boars destined for AI centres (PCVAD)

12.1 Porcine Circovirus Associated Diseases (PCVAD) – diagnostic criteria for infection among a herd¹

- ❖ Criterion 1: Clinical signs among the herd:
 - Wean-to-finish mortality exceeds 8%.
 - Increase in monthly mortality rate equal to or greater than 1.66 standard deviation (SD), when compared to historical data.

- ❖ Criterion 2: Pathological and histopathological lesions:
 - Identification of the typical lesions of PCVAD in at least one pig from the herd that is showing clinical signs (Criterion 1);
 - Testing for typical PCVAD lesions in a minimum of four pigs from the identified population. Two consignments of two piglets less than ten days apart are recommended.

12.2 Evaluation Strategy

A herd becomes suspect when clinical signs meet Criterion 1 (the cause of excess mortality must be explained in a written document from the veterinarian in charge and a laboratory report must confirm the diagnosis).

A herd becomes positive when both Criteria 1 and 2 are met.

¹ The definition is adapted from the definition proposed by the European consortium's research team on PCVAD.

13 Certification of the health status of boars for the centres (Senecavirus A)

13.1 Recommended laboratory technique

- ❖ PCR test for Senecavirus A.

13.2 Evaluation strategy

- ❖ A signed health certificate confirming the absence of clinical signs of disease throughout the growth period received within **two weeks** prior to moving the boars from the farm to AI centres quarantine
- ❖ Verify the absence of the virus with the PCR technique using one of two options:
 - Option A)** Oral fluids with the rope technique in a minimum of four pens;
 - Option B)** Manure samples are taken in a minimum of 4 pens with the possibility to sample the boots of the producer or the walkway.
- ❖ Verification of the absence of Senecavirus A will be confirmed by the PCR test on four samples (which can be pooled by 4)
- ❖ The sampling should be done as close as possible to the shipment of the boars and no more than 30 days prior shipment of the boars in quarantine.

13.3 Interpretation of the PCR tests

Each result is interpreted individually:

- ❖ Interpretation of the test for the Senecavirus A:
 - CT value < 35 ➡ positive sample;
 - CT value ≥ 35 et ≤ 38 ➡ suspicious sample;
 - CT value > 38 ➡ negative sample.

Note: The interpretation of these tests may change depending on the new scientific knowledge that emerges.

13.4 Possible answers

- ❖ All results are negative compared to the PCR test for Senecavirus A:
 - Boars are accepted.
- ❖ Some results are suspicious or positive:
 - Request additional tests on positive samples already in the laboratory:
 - Separate (depool) and repeat the PCR test.
 - Take oral fluids (minimum of 4) or manure samples from four parks (those nearby):
 - Do the PCR test on oral fluids or manure samples.
 - If only one of the samples is positive in relation to any of the tests, no boar is allowed in the centre.

14 Surveillance procedures for boar stud units (PRRS)

14.1 Recommended laboratory tests

- ❖ IDEXX ELISA-X3 test (to detect antibodies)
- ❖ MFIA test (Biovet) or Biocheck test (FMV)
- ❖ PCR test (to detect virus)

14.2 Minimum testing strategy by boar population

- ❖ Once a week:
 - PCR on a pool of ten serums.
- ❖ Once a month (first week of month):
 - ELISA-X3 on five serums.

Note 1 The selected animals are those with health or productivity problems (anorexia, semen quality, fever).

Note 2 Boar stud units may comprise a single building (one population) or several premises (many populations).

14.3 Possible responses

- ❖ All results are negative for both PCR testing and the ELISA-X3 test:
 - continuation of operations.
- ❖ One ELISA-X3 result is positive or suspect:
 - re-test using MFIA test (Biovet) or Biocheck test (FMV) do PCR testing if it has not already been done.
- ❖ One PCR result is positive:
 - immediate alert and possible suspension of operations at the centre (see Emergency procedure in case of PRRS alert in an AIC).

14.4 Monitoring the incidence of health problems potentially linked to PRRS

- ❖ Incidence (number of new cases) of the following health problems is recorded each day:
 - number of boars not eating
 - number of boars producing poor quality semen
 - number of boars coughing or having trouble breathing.

An abnormal increase in the incidence of these problems (twice as much as usual) over a period of three days is cause for an immediate alert (see Emergency procedures in case of a PRRS alert in an AIC).

15 Emergency procedures in case of a PRRS alert in an AIC

15.1 Positive blood test result

The insemination centre that receives a positive blood test result (PCR testing or a positive ELISA-X3 test) must take these two steps within the following 24 hours:

- ❖ Request additional tests be done on the positive serums already at the laboratory:
 - redo the PCR testing or the ELISA-X3 test
 - do MFIA test (Biovet) or Biocheck test (FMV)
- ❖ Do new blood draws from the suspect boars and from the neighbouring boars (the two on the right, the two on the left, the two behind and the two in front):
 - do the ELISA-X3 test on each serum and a PCR on pools of three serums.
- ❖ If the positive result comes from a PCR test, the sale of semen from all boars located in the same building must be suspended until further tests are carried out.

15.2 Increase in health problems

An insemination centre that sees a significant increase in the incidence of health problems potentially related to PRRS must take the following measure within the next 24 hours:

- ❖ Take blood draws from the boars with health problems and from the neighbouring boars (the two on the right, the two on the left, the two behind and the two in front):
 - do the ELISA-X3 test on each serum and a PCR on pools of three serums.

15.3 Possible responses

- ❖ Results of the further tests are negative:
 - continuation of operations
 - a new verification (ELISA-X3 and PCR) of the suspect boars and their neighbours three days after the alert.
- ❖ Results of the further tests are positive:
 - the centre is declared positive
 - suspension of all operations

16 Procedures to follow whenever an AIC is infected with PRRS virus

16.1 Insemination centre is positive

- ❖ Harvest serum from each boar (within less than 24 hours of identifying the problem):
 - do ELISA-X3 test on each serum
 - do PCR testing on the pools of five serums
 - do PCR testing on each serum from the positive pools
 - identify the positive and the suspect boars
 - identify all the farms that received potentially infected semen and alert them to the situation.

Note An insemination centre taking part in PGSCIA that finds itself infected with the PRRS virus is not responsible for the actions to be taken at the client's farm. However, the insemination centre manager may make the following suggestions in the letter to farmers:

- ❖ Advise the farmer/herdsman to inform his consultant veterinarian as soon as possible so measures judged appropriate can be taken.
- ❖ Suggest the recommendation from the CDPQ-designated veterinarians on what to do in case of a PRRS outbreak in an AIC:
 - put any sows that were inseminated with the suspect semen into an isolation (quarantine) unit and wait one month before verifying their health status
or
 - send all sows inseminated with the suspect semen to the slaughterhouse

17 Surveillance procedures for boar stud units (PEDV, TGE and SDCV)

17.1 Recommended laboratory tests

- ❖ PCR test (to detect virus).

17.2 Minimum testing strategy by boar population

- ❖ Once a week:
 - PCR on a pool of five fecal samples per building;

Note 1 The selected animals are those with health or productivity problems (diarrhea, anorexia, semen quality, fever).

Note 2 Boar stud units may comprise a single building (one population) or several premises (many populations).

17.3 Possible responses

- ❖ All results are negative:
 - continuation of operations;
- ❖ One result is positive:
 - immediate alert and possible suspension of operations at the centre (see Emergency procedures in case of a PEDV, TGE or SDCV alert in an AIC).

17.4 Monitoring the incidence of health problems potentially linked to PEDV and SDCV

- ❖ Incidence (number of new cases) of the following health problems is recorded each day:
 - number of boars not eating;
 - number of boars vomiting;
 - number of boars with diarrhea or soft feces.
- ❖ An abnormal increase in the incidence of these problems (twice as much as usual) over a period of two days is cause for an immediate alert (see Emergency procedures in case of a PEDV, TGE or SDCV alert in an AIC).

18 Emergency procedures in case of a PEDV, TGE or SDCV alert in an AIC

18.1 Positive blood test result

The insemination centre that receives a positive test result (positive PCR testing) must take the following two measures within the next 24 hours:

- ❖ Request that additional tests be done on the positive samples already at the laboratory:
 - redo the PCR testing;
- ❖ Take fecal samples or rectal swabs from the suspect boars and from the neighbouring boars (the two on the right, the two on the left, the two behind and the two in front):
 - do the PCR testing on the fecal samples or rectal swabs.
- ❖ The sale of semen from all boars located in the same building must be suspended until further tests are carried out.

18.2 Increase in health problems

An insemination centre that sees a significant increase in the incidence of health problems potentially related to PEDV, TGE or SDCV must take the following measure within the next 24 hours:

- ❖ Take fecal samples or rectal swabs from the boars with health problems and from the neighbouring boars (the two on the right, the two on the left, the two behind and the two in front):
 - do a PCR test on the fecal samples or rectal swabs.

18.3 Possible responses

- ❖ Results of the further tests are negative:
 - continuation of operations;
 - new verification (PCR) of the suspect boars and their neighbours three days after the alert;
- ❖ Results of the further tests are positive:
 - the centre is declared positive;
 - suspension of all operations.

19 Procedures to follow when an AIC is infected with by PEDV, TGE or SDCV virus

19.1 Positive insemination centre

- ❖ Identify all the farms that have received potentially contaminated semen and alert them to the situation.

Note: An insemination centre taking part in PGSCIA that finds itself infected with the PED, TGE or SDCV virus is not responsible for the actions to be taken at the client's farm. However, the insemination centre manager may make the following suggestions in the letter to farmers:

- ❖ Advise the farmer/herdsman to inform his consultant veterinarian as soon as possible so measures judged appropriate can be taken.

20 Microbiological quality control of the semen

20.1 Objectives

- ❖ Sperm survival
- ❖ Bacteriological quality of the diluted semen

20.2 Methods

20.2.1 Bacteriological monitoring of the semen sent to farms

Each week, three to five boar semen samples that have been stored for 48 hours, are selected for bacteriological testing.

Two methods are to be used in the AICs:

1. Rapid non-qualitative method:
 - a) on-site analysis with 3M's Petrifilm[®] technology:
 - One millilitre (1 ml) of semen is inoculated and incubated for 72 hours at 30° C.
 - Bacteria are counted, and the information is recorded in a logbook.
 - A bacterial count of more than 10 colonies per ml in three consecutive samples suggests that there may be a problem. Inform those in charge.
 - b) Alternatively, the on-site analysis can be done with the Compact Dry[®] technology.
2. Precise qualitative method: laboratory analysis by MAPAQ's own laboratory (see Appendix 5) (minimum of five samples per month).

20.2.2 Bacteriological monitoring of the water

The bacteriological quality of the water will be assessed on a regular basis (min. once per month). Water containing more than 10 colonies will be considered inadequate for semen preparation.

Appendix 1 - Registration Form for AI centres

Note: Fill out one form for each boar stud unit

GENERAL INFORMATION	
Company name:	
Boar stud unit name:	
Boar stud unit address:	
Contact name:	
Phone:	
Cell phone:	
Fax:	
E-mail:	
Billing address:	
Number of pig-places:	

Participation fees for the PGSCIA program for 2023-2024 have been set at \$420 per boar stud unit.

The fees per batch of animals assessed by the CDPQ-designated veterinarian vary according to the size of the batch:

- \$70: 10 boars or less
- \$80: 11-20 boars
- \$90: 21-40 boars
- \$100: more than 40 boars

The member certifies that this information is correct and undertakes to comply with the registration requirements of the PGSCIA for 2023-2024.

Owner or Agent

Date

Appendix 2 - Contact Details for the Resource Persons

Head office

Centre de développement du porc du Québec inc.
815 Route Marie-Victorin
Lévis (secteur St-Nicolas) G7A 3S6
Phone : 418 650-2440
Fax : 418 650-1626
E-mail : labo-sante@cdpq.ca

Resources

Marie-Claude Poulin, D.V.M., D.A.
Phone : 418 522-6015
E-mail : marie-claude.poulin@hotmail.co.uk

Claudia Coulombe, Animal health technician
Centre de développement du porc du Québec inc.
Phone : 418 650-2440, poste 4316
E-mail : ccoulombe@cdpq.ca

Christian Klopfenstein, Ph. D., D.V.M,
Centre de développement du porc du Québec inc.
Phone : 418 650-2440, poste 4309
E-mail : cklopfenstein@cdpq.ca

Appendix 3 - CDPQ Health Certificate for Boar Entry into the AIC (D03)

Batch from a PRRS negative farm

Farm name _____ Inventory in finishing unit _____

% Mortality	Months of the year (the last 6 months)					
	_____	_____	_____	_____	_____	_____
Preweaning	_____	_____	_____	_____	_____	_____
Nursery	_____	_____	_____	_____	_____	_____
Finishing	_____	_____	_____	_____	_____	_____

Disease	Clinical signs	Date of last testing	Laboratory			Health Status ¹	
			ELISA	PCR	Others	Positive	Negative
Enzootic pneumonia	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Pleuropneumonia (App)	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
<i>Actinobacillus suis</i>	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Glässer's disease	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
PRRS	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Proliferative enteropathy	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
PCVAD	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Dysentery	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Atrophic rhinitis	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Sarcoptic mange	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
PED	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Delta coronavirus (SDCV)	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
TGE	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Senecavirus A	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Influenza	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Salmonella	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Other significant health problems	_____						
Have your animals been vaccinated?		_____					
If yes, what vaccines were used?		_____					
Date of the last veterinarian's visit (less than 3 months)		_____					

1- Establish the health status based on laboratory results, veterinary visits, slaughterhouse controls or any other diagnostic means.

I, the undersigned, declare that I have forwarded to the veterinarian all the information necessary to assess the health of my herd. Furthermore, in the event of significant changes in the months following the delivery of my animals (boars), I undertake to promptly inform the CDPQ-designated veterinarian.

Owner or Agent

Date

I, the undersigned, certify that the above information is, to the best of my knowledge, complete and accurate.

Veterinarian

Date

Appendix 4 - Laboratories

Serological tests for *Actinobacillus pleuropneumoniae*

- The animal health services laboratories of the Service de diagnostic de la Faculté de médecine vétérinaire (FMV) of the Université de Montréal

Serological tests for porcine reproductive and respiratory syndrome (PRRS)

- Biovet laboratory in Saint-Hyacinthe
- The animal health services laboratories of the Service de diagnostic de la Faculté de médecine vétérinaire (FMV) of the Université de Montréal

Tests for porcine epidemic diarrhea (PEDV), transmissible gastroenteritis (TGE) and diarrhea caused by porcine delta coronavirus (SDCV)

- Biovet laboratory in Saint-Hyacinthe
- The animal health services laboratories of the Service de diagnostic de la Faculté de médecine vétérinaire (FMV) of the Université de Montréal

Tests for Senecavirus A

- Biovet laboratory in Saint-Hyacinthe
- The animal health services laboratories of the Service de diagnostic de la Faculté de médecine vétérinaire (FMV) of the Université de Montréal

Microbiological analysis (semen) and autopsies

- The animal pathology laboratory of the ministère de l'Agriculture, des Pêcheries et de l'Alimentation du Québec (MAPAQ)
- The animal health services laboratories of the Service de diagnostic de la Faculté de médecine vétérinaire (FMV) of the Université de Montréal

Appendix 5 – Sampling guide for boar purchases (D02)



Sampling guide for boar purchases

Client :	CDPQ-PGSCIA	
Address :	815 Rte Marie-Victorin Lévis, (secteur Saint-Nicolas) G7A 3S6	Phone : 418-650-2440 E-mail : labo-sante@cdpq.ca

Instructions to the CFIA veterinarian

- Please send a portion of the sample (serum, feces or oral fluid) from each boar or each pen to the laboratory to Biovet or FMV for further analysis.
- Complete the official laboratory form and send a copy of the results to the to the CDPQ by email at labo-sante@cdpq.ca
- The letters¹ identifying the company, the tattoo number of each boar or the number of the pen must be written on the containers.
- The samples must be always kept cool (4°C).
- We also require you to check off the tests App serotypes 1 and 5 if they were asked for. The breeder must be informed when CDPQ asks for these tests. When in doubt, contact the CDPQ.

Mandatory tests

- PRRS IDEXX ELISA-X3 on all serums;
- PRRS PCR on 5 serums (select the samples with the highest S/P ratios (IDEXX ELISA-X3), including positive or suspect serums), request to be tested in 1 pool of 5;
- PED, TGE and SDCV on **1 pool of 4 pens** (environmental sampling or oral fluids);
- Senecavirus A PCR on **1 pool of 4 pens** (environmental sampling or oral fluids).

Test optionnel

- App 1 ELISA et App 5 ELISA

¹ The site code matches the pure breed letters.



Centre de développement du porc du Québec inc.

815 Route Marie-Victorin

Lévis (Secteur St-Nicolas) G7A 3S6

 418 650-2440  418 650-1626

cdpq@cdpq.ca

www.cdpq.ca

 @cdpqinc

